# Claudio Ferfuia, Maurizio Turi and Gian Paolo Vannozzi\* Variability of Seed Fatty Acid Composition to Growing Degree-Days in High Oleic Acid Sunflower Genotypes

Abstract: High temperature enhances the oleic acid content in the oil of normal cultivars but conflicting results are reported on temperature effects on oleic acid content in HO cultivars: either no effect or an increase in oleic acid content with temperature. To investigate the effects of temperature on HO genotypes under natural field conditions, a three-year field trial was conducted using two sowing dates and three HO genotypes (two inbred lines and one hybrid). To compare our results with previous works, growing degree-days (GDD) were computed (base temperature  $=6^{\circ}$ C). GDD accumulated during the "flowering – 25 days after flowering" period influenced fatty acid composition of seed. Oleic and linoleic acid contents were affected by accumulated GDD in two HO genotypes (one inbred line and the hybrid). There was an increase of about 3% in oleic acid content as response to more high GDD accumulated. Their content was not modified by GDD in the other inbred line. There was a genotype  $\times$  environment interaction that we suppose depending on modifier genes. These genetic factors affected oleic acid content. This indicated the importance of breeding targeted to select hybrids with a stable oleic acid content and higher than 90%. Saturated fatty acids (palmitic and stearic) were also influenced by temperature, and there was genetic variability among genotypes.

Keywords: Helianthus annuus, planting date, seed oil content, growing degreedays, fatty acids composition, high oleic hybrids

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## Introduction

Sunflower (Helianthus annuus L.) is one of the most important oilseed crops in the world. Sunflower oil has a wide range of applications, both in the food industry (e.g. margarine production) and as raw material for non-food application (biofuels, oleochemicals). The quality of sunflower seed oil is determined by the content and ratio of fatty acids. It contains both saturated (palmitic and stearic acid) and unsaturated fatty acids (up to 90–95%), either monounsaturated (oleic acid) or polyunsaturated (mostly linoleic and traces of other minor fatty acids). Depending on unsaturated fatty acid composition, sunflower cultivars can be divided into traditional or normal or low oleic  $(LO - < 40\%$  oleic acid) types, with an oil composition characterized by a majority of linoleic acid, mid-oleic  $(MO - 65-75%)$  oleic acid) and high oleic  $(HO - >80\%$  oleic acid).

Previous research has demonstrated that the fatty acid composition of sunflower oil depends on genotypes (LO, MO and HO) and environmental conditions during seed-filling phase (Harris et al., 1978; Champolivier and Merrien, 1996; Roche et al., 2006; Izquierdo and Aguirrezábal, 2008). HO sunflower was obtained by chemical mutagenesis with dimethyl sulfate (DMS) of LO sunflowers (Soldatov, 1976). HO genotypes cumulate the Pervenets mutation effect and other independent factors acting on oleic acid content such as modifier genes (Miller et al., 1987; Fernandez et al., 1999; Velasco et al., 2000; Lacombe et al., 2004). Modifier genes are minor genes having no known effect except to intensify or diminish the expression of a major gene (Briggs and Knowles, 1967). Some genetic variation for oleic acid content among HO genotypes depends on these independent genetic factors (genetic background of parental lines) from Pervenets mutation, and they fixed the genetic oleic acid content potential of HO genotypes. Phenotypic expression of some of these modifiers can be modified by temperature, and some of them have no phenotypic expression at higher temperatures (Velasco et al., 2000).

It has long been known that temperature is the main environmental factor affecting the fatty acid composition in the oil of LO sunflower (Canvin, 1965), mainly regulating the ratio of oleic and linoleic acids. High temperature enhances the oleic acid content of normal cultivars but conflicting results are reported about temperature effects on oleic acid content of high oleic acid cultivars: either no effect (Lagravére et al., 2000) or an increase in oleic acid content with temperature (Champolivier and Merrien, 1996; Triboï-Blondel et al., 2000; Izquierdo and Aguirrezábal, 2008). These differences could be related to differences in the studied hybrids as well as their genetic backgrounds.

Lagravére et al. (2000) suggested that hybrids with low oleic acid potential could be more sensitive to environmental conditions such as temperature, while hybrids with a higher genetic oleic acid content potential were insensitive to temperature conditions. Under natural field conditions, the effect of temperature on HO genotypes through a delay in sowing has not been extensively studied. Connor and Sadras (1992) reported that sowing date influences the fatty acid composition by modifying the ontogenesis. The same variation is observed for oleic hybrids (Flagella et al., 2002; Roche et al., 2004, 2006) but the variation between dates of sowing was less (a variation in oleic acid content of about 2%).

Another topic of interest is quantifying the effects of temperature on oleic acid content at different stages in the seed-filling phase in sunflower and thus selecting the critical period. Critical periods are those when the sensitivity to an environmental variable is highest. Izquierdo et al. (2002, 2006) and Izquierdo and Aguirrezábal (2008) found increments in oleic percentage with higher night temperatures applied during early stages of grain development (100–300 degree days after flowering, base temperature 6°C), with standard genotypes showing the greatest change and high oleic hybrids the least. Roche et al. (2006) hypothesized that changes in the level of oleic acid in seeds are modulated by the mean temperature during the flowering period and also by the temperature sums of all phases. It is not known whether the period in which temperature has maximum effect on fatty acid composition differs among HO genotypes.

The first issue, therefore, in selecting HO hybrids with a very high content in oleic acid content  $($ >94%) is to avoid the effects of modifier on oleic acid content, or in other word the interaction genotype by environment, to maximize its content. The second issue is concerning the ratio oleic–saturated fatty acid content. The saturated fatty acids, palmitic and stearic, are less influenced by environmental conditions than unsaturated fatty acids with small variations between years and locations (Lajara et al., 1990; Izquierdo et al., 2002, 2006; Vick et al., 2004). However, to select HO hybrids with a high and stable oleic acid content, it is important to detect all factors that may reduce or increase its concentration in the seeds.

In this study, we compared two HO inbred lines, the first with a high genetic oleic acid content potential (R978) while the second had a low potential in oleic acid content (342mt) and their hybrid. This work aimed at assessing the grain fatty acid composition of three HO sunflower genotypes in response to growing degree-days (GDD) accumulation and to study variability in seed fatty acid composition among HO genotypes as affected by temperature.

## Materials and methods

#### Plant material

The sunflower (Helianthus annuus L.) seeds used in this work were from the inbred lines 342mt and R978, both high oleic inbred lines, and from their hybrid  $342 \times R$ 978. Line  $342$ mt is a selection, made at the University of Udine, derived by Ha 342 USDA, and it is a male sterility maintainer with a single head. Line R978, selected by the University of Udine, is a fully branched type and is a male fertility restorer. The lines are maintained at the University of Udine, and they have been used in a high oleic breeding program for 20 years. The behaviors, concerning seed fatty acids composition, of these HO lines are well established in the environmental conditions of North-East Italy. R978 inbred line has a high genetic potential in oleic acid content while 342 has a low potential.

#### Field trials

Inbred lines and hybrid were grown in 2009–2011 on the experimental farm of the University of Udine, Azienda Agraria Universitaria "A. Servadei" (46°04ʹN, 13°22ʹE, 109 m a.s.l.), in North-East Italy. The experiment was designed as a completely randomized block scheme, with three replications, using two sowing dates. Plot size was 5 m  $\times$  2 m. The seeds were sown with a spacing of 0.75 m between rows. Plants were thinned after seedling emergence from 10 to 7.5 plants m<sup>-2</sup>. Nitrogen was applied at 100 kg ha<sup>-1</sup>. Weeds and diseases were controlled, and regular watering throughout the experiment ensured that plants were not subjected to water deficit during the entire growing period. At the R4 stage, all plants were covered with paper bags (Schneiter and Miller, 1981) to prevent cross-fertilization. Five plants were studied per plot.

Meteorological data (Table 1) for the experimental period were recorded at a weather station (Udine – S. Osvaldo Station; Osmer- FVG Region Meteorological Service) located 200 m away from the field site.

#### Sampling

Harvesting was done when all plants in a given treatment reached physiological maturity (PM), R9 phase (Schneiter and Miller, 1981). Seeds from outer rings

	2009			2010			2011		
	$T_{\min}$ (°C)	$T_{\rm max}$ (°C)	Rainfall (mm)	$T_{min}$ (°C)	$T_{\rm max}$ (°C)	Rainfall (mm)	$T_{\min}$ (°C)	$T_{\rm max}$ (°C)	Rainfall (mm)
April	9.2	20.6	131.5	6.7	19.6	75.1	7.9	22.1	18.1
May	13.1	26.1	28.0	11.7	21.6	230.2	11.5	26.3	85.2
June	15.2	26.8	104.2	15.5	27.1	68.7	15.6	26.7	185.1
July	17.1	29.7	104.5	18.1	30.6	143.7	15.8	28.1	148.4
August	18.2	31.8	66.2	16.2	28.0	122.1	16.9	31.3	23.3
September	14.5	26.9	145.6	12.2	23.4	264.8	14.6	29.3	83.9
Mean	14.5	27.0		13.4	25.1		13.7	27.3	
Total			580.0			904.6			544.0

Table 1: Meteorological data for the field site during sunflower growth in 2009–2011.

were separated for fatty acid and oil content determination and analyzed. Seeds were dried in an oven at 60°C for 48 h.

To find the real PM, and thus to investigate the relationship between fatty acids contents and temperature, three plants per plots were sampled twice per week over a period of 5 weeks, commencing 10 or 13 days after flowering (daf) for a total of 10 samplings. At each sampling ten seeds per plant from each of three plants were taken from the outer region of the capitulum (first six rings). Seeds were dried in an oven at 60°C for 48 h. Plants reached true PM, when the seed weight became constant after three successive samplings.

#### Fatty acid analysis

Five plants per plot and 100 seeds per plant from the outer rings (1–6) were collected and dehulled. Kernels were ground to a fine powder using a coffee grinder. Two hundred milligrams of kernel powder was weighed to perform fatty acid analysis. Lipids were extracted in  $n$ -hexane. Fatty acids were converted in Fatty Acid Methyl Esters (FAMEs) by transesterification with a methanolic potassium hydroxide solution (2 N). FAMEs composition was determined by gas chromatography, and every fatty acid was expressed as a percentage of the total fatty acids detected in the oil. The gas chromatograph, equipped with a Flame Ionization Detector (FID) and a split–splitless injector, was fitted with a 60 m HP-88 capillary column (Agilent Technologies, USA). Helium was used as carrier gas, and the injector, detector and oven temperatures were 280, 250 and 200°C, respectively. Five microliters of sample was injected in split mode. Different FAMEs were identified by comparison with known standards.

#### Oil content analysis

Whole seeds, 10 g, before fatty acid determination, were analyzed for oil content by Nuclear Magnetic Resonance (NMR Oxford Instruments –4000; FGIS-USDA, 2009).

#### Growing degree-days accumulation

To study the environmental effects on fatty acids and to compare our results with previous works, data from years and date of sowing were divided according to GDD accumulated. The accumulation of GDD, above a base temperature of  $6^{\circ}C(T_b)$ , was calculated using the following formula:

$$
\text{GDD} = \Sigma[(T_{\text{max}} + T_{\text{min}})/2] - T_{\text{b}}
$$

where  $T_{\text{max}}$  and  $T_{\text{min}}$  are the daily maximum and minimum temperatures, respectively, in °C. GPM is calculated from last flowering to PM and G25 from last flowering to 25 daf. Two GPM groups were created: A and B. Group A contained inbred lines at 500 GGD and the hybrid at 600 GGD. Group B was formed by inbred lines at 600 GGD and the hybrid at 700 GGD. G25 was also separated into two groups: the first one was constituted by plants that had accumulated 400 GGD and the second by plants that had accumulated 500 GGD.

### Statistical analysis

Statistical analysis was performed using R version 2.15.0 (R Development Core Team, 2012). Shapiro–Wilk normality test was performed to test normality condition. A two-way ANalysis Of VAriance (ANOVA) was performed as fixed-effect model with GDD and genotypes. Significance of each source of variation was evaluated by  $F$ -test. When the  $F$ -ratio revealed significant differences, means were compared by the least significant difference (LSD) at  $p \le 0.05$ .

# Results

Duration of the growing period (emergence – PM) and relative phases are reported in Table 2. PM, expressed as daf when seed weight was constant, was reached some days before R9 phase (Schneiter and Miller, 1981). In inbred lines the difference between PM and R9 was smaller (about 4 days before than R9



Table 2: Date of sowing, cycle duration and emergence to flowering, and end of flowering (F)-PM phase duration for each genotype.

Notes: <sup>a</sup>PM - R9 according to Schneiter and Miller (1981); <sup>b</sup>PM as seed constant weight.

phase) than in the hybrid (about 10 days before than R9 phase). Thus, the duration of end of flowering to PM phase was calculated on true PM expressed as daf when seed weight became constant.

The GDD accumulated from last flowering to PM (GPM) were  $618 \pm 49$ (standard deviation) for the year 2009,  $540 \pm 49$  and  $630 \pm 38$  GDD for the years 2010 and 2011, respectively. Average GPM accumulated by inbred lines was 567 °C days  $\pm$  67 while the hybrid accumulated 652 °C days  $\pm$  53. The GDD accumulated from last flowering to 25 daf (G25) was  $439 \pm 33$ ,  $426 \pm 22$  and  $405 \pm 23$  G25 for 2009, 2010 and 2011, respectively. Average G25 accumulated by inbred line 342mt was  $436 \pm 34$ ,  $420 \pm 28$  by inbred line R978 and  $414 \pm 23$  by the hybrid. All tested genotypes accumulated the same GPM and G25 on average.

Oil and stearic acid contents were influenced by the interaction genotype  $\times$  GPM accumulated from flowering to PM (Tables 3 and 4). The composition of seed fatty acids (palmitic acid and unsaturated oleic and linoleic acids) was influenced by G25 accumulated (Table 5) from last flowering to 25 daf. Line 342mt and hybrid showed a different fatty acid composition at



Table 3: ANOVA (mean square) for the main fatty acids and seed oil content.

Notes: GPM are the GDD accumulated from end of flowering to PM (daf when seed weight becomes constant). \*, \*\* and \*\*\* = Significant at the  $p < 0.05$ , 0.01 and 0.001 levels, respectively.  $ns = not$  significant.

Genotype GPM Stearic acid % Oil % 342mt 500 2.7ab 44.6bc 600 3.3a 40.6c R978 500 500 2.7ab 42.2c 600 2.1b 47.9ab Hybrid 500 2.0b 51.2a 600 3.0a 48.1ab

Table 4: Stearic acid and oil content as affected by GPM (GDD accumulated from last flowering to PM) accumulated in the tested genotypes.

Note: Means followed by the same letter are not significantly different (LSD at the 5% level).





Notes: G25 are the GDD accumulated from end of flowering to 25 daf. \*, \*\* and \*\*\* = Significant at the  $p < 0.05$ , 0.01 and 0.001 levels, respectively; ns = not significant.

different levels of G25 while line R978 had the same fatty acid composition at every level of G25. Thus, temperature did not modify fatty acid composition in line R978, but it depended only on genotype.

Palmitic acid content was 4.20% on average, and it was the main saturated fatty acid in tested HO genotypes. The two high oleic inbred lines tested and

Genotype	G <sub>25</sub>	Palmitic acid %	Oleic acid %	Linoleic acid %
342mt	400	3.8 <sub>bc</sub>	84.8c	8.1a
	500	5.4a	88.2b	3.8 <sub>b</sub>
R978	400	3.3c	92.0a	2.5 <sub>b</sub>
	500	4.9ab	90.8ab	1.4c
Hybrid	400	4.0c	89.9b	3.5 <sub>b</sub>
	500	3.6 <sub>b</sub> c	92.9a	1.2c

**Table 6:** Effects of interaction genotype  $\times$  G25 (GDD accumulated from last flowering to 25 daf) on palmitic, oleic and linoleic acid contents.

Note: Means followed by the same letter are not significantly different (LSD at the 5% level).

their hybrid had different palmitic acid contents (Table 6). The seeds of inbred lines had an increase in palmitic acid content with G25B. The highest value was reached by line 342mt with G25B (5.37%), and the lowest by R978 inbred line with G25A (3.28%). We reported a positive correlation between mean daily temperature of the F-PM phase and palmitic acid content only in inbred line 342mt (Figure 1), while there was no any correlation between temperature and palmitic acid content in inbred line R978. The palmitic acid content in the hybrid was within the parental range, and it did not show any variation linked to environmental condition.

The stearic acid content of the inbred lines was unaffected by heat summation, whereas it increased with GPM2 in the hybrid. The hybrid had a stearic



Figure 1: Correlation between mean air temperature and seed palmitic acid content in inbred line 342mt during F-PM phase (p-value 0.015).

acid content within the parental range and showed the same interaction genotype  $\times$  GPM as its female parental line. The hybrid showed an increase of about 1% in stearic acid content when GPM increased by 100°C days. Inbred lines had the same stearic acid content through the GPM levels. At the level A of GPM accumulated, there was no significant difference among tested genotypes. At the level B of accumulated, inbred line R978 had the lowest stearic acid content (2.0%).

Small but significant variations in oleic acid content were manifested with different levels of G25 and with a difference among genotypes. ANOVA showed that there was a relative prominence of genotype effects compared to G25 (Table 5). The two high oleic inbred lines tested differed about their oleic acid content. Inbred line 342mt showed a lower oleic acid content on average than R978 inbred line and their hybrid (Table 6).

Oleic and linoleic acids were negatively correlated. There was a difference between high oleic inbred lines (Figure 2). Line 342mt showed a strong negative oleic–linoleic relationship ( $p$ -value < 0.001), indicating that increasing one point in oleic corresponded to a decrease of one point in linoleic acid. The correlation coefficient between oleic and linoleic acids was not significant in R978 (Table 7). The hybrid showed a strong negative oleic–linoleic relationship and had an average value of the regression parameter among parental lines (Figure 2). This suggests an additive effect of modifier genes on oleic acid content.



Figure 2: Relationship between oleic and linoleic acids in inbred lines and in the hybrid.

Linoleic acid content in seeds was significantly affected by G25 only in two genotypes: 342mt and the hybrid (Table 6). As for oleic acid content, linoleic acid content in inbred line R978 did not show any response to the G25 tested.



Table 7: Correlation coefficients between seed oil content and concentration of fatty acids in the three high oleic genotypes tested  $(n=18)$ .

Notes: \*, \*\* and \*\*\* = Significant at the  $p < 0.05$ , 0.01 and 0.001 levels, respectively.

Line 342mt and the hybrid showed a reduction in linoleic acid when heat summation dropped of 100°C days.

Oil content varied from 40 to 51%. ANOVA showed that oil content was influenced by genotype effects and the interaction genotype  $\times$  GPM (Table 3). The hybrid showed a higher seed oil content than its parental lines. 342mt and the hybrid did not show any significant variation with the GPM tested while R978 showed an increase in seed oil content when GPM increases from 500 to 600°C days. There was no significant relationship between seed oil content and oleic acid content in any of the genotypes (Table 7). In inbred line 342mt, oil content was negatively correlated with stearic acid content while oil concentration in the hybrid was positively correlated with linoleic acid concentration.

## **Discussion**

The GDD summation (flowering  $-$  PM) reported in this study compared with 699–836 (mid-flowering – PM) reported by Robertson and Green (1981), when these data are converted to a  $T<sub>b</sub>$  of 6°C, and with the GDD summation (449–948°C days; R6–R9 phase) reported by Roche et al. (2006). Different sunflower genotypes

grown in diverse environments required a similar cumulative heat summation from flowering through PM.

Fatty acids composition was affected by the cumulative heat summation between flowering and 25 daf (G25). This result is in agreement with Izquierdo et al. (2002) who reported that the sensitive period for modifications in fatty acid composition was 0–400°C daf.

#### Effect of GDD on saturated fatty acids content

The oleic genotypes had lower palmitic acid percentages compared with the standard sunflower types as reported in other works (Lajara et al., 1990; Roche et al., 2004; Anastasi et al., 2010). High oleic genotypes with high oleic acid potentials had the lowest palmitic acid contents. Thus, increasing in oleic acid content to select high-HO genotypes could be achieved only through a reduction in palmitic acid content. High levels of saturated fat consumption are correlated with increased risk of coronary heart disease. Despite sunflower oil already having a high level of unsaturated fatty acids, it is still desirable to reduce its saturated fatty acid level for producing healthier oil.

Palmitic acid concentration increases with G25B only in the inbred lines. Several environmental factors like water availability (Roche et al., 2006; Jalilian et al., 2011), temperature (Rondanini et al., 2003; Izquierdo and Aguirrezábal, 2008) and nitrogen (Zheljazkov et al., 2009) could alter saturated fatty acid content in sunflower. In our experiment, temperature was the most variable environmental factor. The temperature modified the palmitic acid concentration of the oil in some high oleic hybrids (Izquierdo and Aguirrezabal, 2008). We found a positive correlation between mean daily temperature of the F-PM phase and palmitic acid content only in inbred line 342mt (Figure 1) while there was no any correlation between temperature and palmitic acid concentration in inbred line R978. Interestingly, oil seed percentage and palmitic acid were positively correlated in inbred line R978 (Table 7). The seed oil content increased with GPM2 (Table 6). The rise in palmitic acid content was related to an increase in seed oil content and not to a rise in air temperature per se. Thus, heat summation did not modify palmitic acid percentage in this line, but oil content.

Stearic acid concentration was reduced in tested HO genotypes with respect to LO sunflower. Stearic acid concentration was associated with an increase in heat sum only in the hybrid whereas the concentration was unaffected in the inbred lines. The difference could be associated with genotype (e.g. Roche *et al.*, 2006).

As hypothesis, differences among genotypes could be related to isozymes of stearate desaturase (Fernandez-Moya et al., 2003). About the isozymes, at  $GPM<sub>A</sub>$ , mean air temperature was lower than at  $GPM_B$  (21.9°C and 23.6°C, respectively). In soybean, Byfield and Upchurch (2007) found that decreased SAD transcript accumulation at warmer temperature was positively associated with a significantly increased level of stearic acid but only in a high stearic mutant line. Conversely, in a soybean genotype, the stearic acid percentage was negatively related to daily mean temperature during grain-filling period (Zuil *et al.*, 2012). The effect of temperature on stearic acid content is unclear and seems to be genotype-specific. Considering that variation in stearic acid amount was small, the response to GPM could be related to a "long-term" effect of temperature on stearic acid content.

Interestingly, the two saturated fatty acids displayed an opposite interaction with heat summation between parental lines and hybrid. Palmitic acid concentration was affected by temperature in inbred lines and not in the hybrid while these genotypes showed an opposite behavior on stearic acid concentration. The inheritance and the environmental effects on saturated fatty acids content are composite. Palmitic and stearic acids are both quantitatively inherited. Genetic background and environment also play significant roles in palmitic and stearic acid inheritance (Roche et al., 2006). In the literature, there are only a few works on inheritance of saturated fatty acids content, and the trait appears to be complex and multigenic (Vick et al., 2004). Further studies are needed on inheritance of saturated fatty acids in HO genotypes.

#### Effect of GDD on C18 unsaturated fatty acids content

Oleic acid contents were relatively stable in the high oleic genotypes tested. These data are in agreement with Lagravére *et al.* (2004) and Roche *et al.* (2004). Oleic acid content was insensitive to the heat sum in the line R978. On the other hand, line 342mt and the hybrid, that they had a different oleic acid content in their seeds (86.5 and 91.4%, respectively), showed the same interaction genotype  $\times$  G25 with an increase of 3% in oleic acid content as heat summation increased from 400 to 500°C day. This indicates the presence of a genotype  $\times$ environment interaction on oleic acid concentration. Flagella et al. (2002) reported a similar variation in oleic acid content (about 2%) in other high oleic sunflower hybrids as affected by date of sowing. Oleic acid content is modified by temperature in some genotypes, thus high oleic inbred lines with different origins are not equivalent for high oleic trait. The basis for differences between high oleic and normal sunflower genotypes is a differential activity of the

enzyme Δ12-desaturase, which catalyzes the desaturation of oleic acid to linoleic acid (Garcés and Mancha, 1991; Kabbaj et al., 1996). High oleic mutants had substantially lower FAD2-1 desaturase gene transcript accumulation than sunflower standard type (Kabbaj et al., 1996; Hongtrakul et al., 1998). The high oleic trait was controlled by at least three loci: oleHL, a suppressor locus and modifier loci (Lacombe *et al.*, 2004). We suppose that the inbred lines and thus the hybrid tested were homozygous at the Pervernets locus. Thus, observed differences can be related only to modifier genes (Miller et al., 1987; Velasco et al., 2000; Varès *et al., 2002; Lacombe et al., 2004).* The genotype  $\times$  environment interaction was caused by modifier genes. The observed response to temperature in inbred line 342mt and the hybrid could be related to a residual activity of the Δ12-desaturase (mediated by modifier) or to some genes that have no phenotypic expression at higher temperatures (Velasco et al., 2000). From a plant breeder's point of view, knowledge of the  $G \times E$  interaction facilitates the efficient use of appropriate breeding and selection procedures. A hybrid that is stable in different growing conditions is preferred. The difference among genotypes in their linoleic acid content was related to modifier genes, as for oleic acid, because there is a close relationship between oleic and linoleic acid synthesis.

#### Seed oil content

Seed oil concentration in this study was generally high and similar to previous reports (Roche et al., 2004; Anastasi et al., 2010).

Results suggest that sunflower seed oil concentration depends on genotype, but it may be expressed differentially under different environmental conditions because seed oil concentration of sunflower is sensitive to environmental conditions during the grain-filling period (Connor and Hall, 1997). No relationship was found between seed oil content and temperature or rainfall. Seed oil content may therefore be affected by other variables not studied in this work. Differences among genotypes in response to GDD could be related to a different oil accumulation pattern (Mantese *et al.*, 2006) or due to the fact that R978 is a fully branched type. Thus, it would be possible to suppose a different interaction between seed characteristic (embryo oil concentration, pericarp/kernel ratio, etc.) and environment.

### Seed oil content and fatty acid relationship

In high oleic hybrids a negative correlation between stearic acid and oil content was reported by Van der Merwe *et al.* (2012). In inbred line R978, oil content was

positively correlated with palmitic acid content. This result is not in agreement with Velasco *et al.* (2007) who reported a negative correlation between oil and palmitic acid contents. In the hybrid, oil content was positively correlated with linoleic acid. Further studies are needed on oil content and its relationship with fatty acids in high oleic mutants.

With regard to the ratio of palmitic to oleic acid, a negative and significant correlation was observed in line R978 and in the hybrid, and these results are in agreement with data by Champolivier and Merrien (1996), Roche et al. (2004) and Izquierdo et al. (2006), collected for oleic hybrids and for high stearic mutants (Velasco et al., 2007). Flagella et al. (2002) showed that an increase in palmitic acid in sunflower is accompanied by a decrease in both oleic and stearic acids. Studies on soybean (Rebetzke et al., 1996), peanut (Andersen and Gorbet, 2002), sesame (Were *et al.*, 2006) and winter oilseed rape (Möllers and Schierholt, 2002) also revealed strong inverse relationships between palmitic and oleic acids. Line 342mt did not show any significant correlation between palmitic and oleic acids, but we found an inverse significant correlation between palmitic and linoleic acids. This could be associated with the relationship between palmitic acid and temperature in this genotype.

Oleic and linoleic acids were negatively correlated. There was a difference between high oleic inbred lines (Figure 2). Line 342mt showed a strong negative oleic–linoleic relationship (*p*-value  $\leq$  0.001), indicating that increasing one point in oleic corresponded to a decrease of one point in linoleic acid. The correlation coefficient between oleic and linoleic acids was not significant in R978 (Table 7). The hybrid showed a strong negative oleic–linoleic relationship and had an average value of the regression parameter between parental lines (Figure 2). This suggests an additive effect of modifier genes on oleic acid content.

We suppose that the linoleic acid level reached by R978 was a physiological threshold for this genotype. We speculate that in R978 only the constitutive desaturase system (FAD2-2 and FAD2-3) present in the whole plant (Martìnez-Rivas et al., 2001) could be active and responsible for low linoleic acid synthesis (physiological threshold) while seed-specific desaturase is fully inactive. The total elimination of linoleic acid from the seed oil by conventional breeding is probably impossible. The negative relationship between oleic acid content and saturated fatty acids suggested that increasing the oleic acid content over 93–95% is possible only with a reduction in saturated fatty acids content. In line 342mt and in the hybrid, linoleic acid synthesis is probably due to a residual activity of the achene-specific desaturase system (Lagravére et al., 2004). This residual activity is regulated by environmental condition.

## Conclusions

Fatty acid composition in high oleic sunflower depended mainly on genotype, but environmental conditions can affect the 90% threshold of oleic acid content. Seed fatty acid composition was mainly influenced by GDD accumulated from end of flowering to 25 daf (G25). At the highest level of G25 accumulated, there was an increase in palmitic and oleic acid contents, whereas linoleic acid content decreased. Oleic acid content was affected by temperature in two of three high oleic genotypes. This result suggests that temperature could be acting on different biological processes in high oleic genotypes. Further studies are needed.

Saturated fatty acids content was affected by GDD with a different response among genotypes. To obtain hybrids with an oleic acid content higher than 93–95% it is necessary to select inbred lines against saturated fatty acid content because the total elimination of linoleic acid from the seed oil by conventional breeding is probably impossible. Further studies are therefore needed on saturated fatty acids content in high oleic genotypes.

## References

- Anastasi, U., Santonoceto, C., Giuffrè, A., Sortino, O., Gresta, F., Abbate, V., 2010. Yield performance and grain lipid composition of standard and oleic sunflower as affected by water supply. Field Crops Research 119: 145–153.
- Andersen, C., Gorbet, D., 2002. Influence of year and planting date on fatty acid chemistry of high oleic acid and normal peanut genotypes. Journal of Agricultural and Food Chemistry 50: 1298–1305.
- Briggs, F.N., Knowles, P.F., 1967. Introduction to Plant Breeding. Reinhold, New York, USA.

Byfield, G.E., Upchurch, R.G., 2007. Effect of temperature on Delta-9 Stearoyl-ACP and Microsomal Omega-6 Desaturase gene expression and fatty acid content in developing soybean seeds. Crop Science 47: 1698–1704.

- Canvin, D., 1965. The effect of temperature on the oil content and fatty acid composition of the oils from several seed crops. Can. J. Bot., 43: 63–69.
- Champolivier, L., Merrien, A., 1996. Evolution de la teneur en huile et de sa composition en acides gras chez deux variétés de tournesol (oléique ou non) sous l'effet de températures différentes pendant la maturation des graines. OCL 3: 140–144.
- Connor, D., Sadras, V., 1992. Physiology of yield expression in sunflower. Field Crops Research 30: 333–389.
- Connor, D.J., Hall, A.J., 1997. Sunflower physiology. In: Schneiter, A.A. (ed) Sunflower Technology and Production, ASA, CSSA, SSSA, Madison, WI, pp. 113–182.
- Fernandez, H., Baldini, M., Olivieri, A.M., 1999. Inheritance of high oleic acid content in sunflower oil. Journal of Genetics and Breeding 53: 99–103.
- Fernández-Moya, V., Martínez-Force, E., Garcés, R., 2003. Temperature-related non-homogeneous fatty acid desaturation in sunflower (Helianthus annuus L.) seeds. Planta 216: 834–840.
- FGIS-USDA, 2009. Nuclear Magnetic Resonance (NMR) Handbook, FGIS-USDA, United States Department of Agriculture, Grain Inspection, Packers and Stockyards Administration.
- Flagella, Z., Rotunno, T., Tarantino, E., Di Caterina, R., De Caro, A., 2002. Changes in seed yield and oil fatty acid composition of high oleic sunflower (Helianthus annuus L.) hybrids in relation to the sowing date and the water regime. European Journal of Agronomy 17: 221–230.
- Garcés, R., Mancha, M., 1991. In vitro oleate desaturase in developing sunflower seeds. Phytochemistry 30: 2127–2130.
- Harris, H., McWilliam, J., Mason, W., 1978. Influence of temperature on oil content and composition of sunflower seed. Australian Journal of Agricultural Research 29: 1203–1212.
- Hongtrakul, V., Slabaugh, M.B., Knapp, S.J., 1998. A seed specific Δ-12 Oleate Desaturase gene is duplicated, rearranged, and weakly expressed in high oleic acid sunflower lines. Crop Science 38: 1245–1249.
- Izquierdo, N., Aguirrezábal, L.A.N., 2008. Genetic variability in the response of fatty acid composition to minimum night temperature during grain filling in sunflower. Field Crops Research 106: 116–125.
- Izquierdo, N., Aguirrezábal, L.A.N., Andrade, F.H., Cantarero, M.G., 2006. Modeling the response of fatty acid composition to temperature in a traditional sunflower hybrid. Agronomy Journal 98: 451–461.
- Izquierdo, N., Aguirrezábal, L.A.N., Andrade, F.H., Pereyra, V., 2002. Night temperature affects fatty acid composition in sunflower oil depending on the hybrid and the phenological stage. Field Crops Research 77: 115–126.
- Jalilian, J., Modarres-Sanavya, S.A.M., Saberalia, S.F., Sadat-Asilanb, K., 2011. Effects of the combination of beneficial microbes and nitrogen on sunflower seed yields and seed quality traits under different irrigation regimes. Field Crops Research 127: 26–34.
- Kabbaj, A., Abbott, A., Bervillé, A., 1996. Expression of stearate, oleate and linoleate desaturase genes in sunflower with normal and high-oleic contents. In: Proceedings of the 14th Intern. Sunf. Conf. Beijing, China, pp. 60–65.
- Lacombe, S., Kaan, F., Griveau, Y., Bervillé, A., 2004. The Pervenets high oleic mutation: methodological studies. Helia 27: 41–54.
- Lagravère, T., Champolivier, L., Lacombe, S., Kleiber, D., Bervillé, A., Dayde, J., 2000. Effects of temperature variations on fatty acid composition in oleic sunflower oil (Helianthus annuus L.) hybrids. In Proceedings of the 15th Intern. Sunf. Conf. Toulouse, pp. 73–78.
- Lagravère, T., Kleiber, D., Surel, O., Calmon, A., Bervillé, A., Dayde, J., 2004. Comparison of fatty acid metabolism of two oleic and one conventional sunflower hybrids: A new hypothesis. Journal of Agronomy and Crop Science 190: 223–229.
- Lajara, J.R., Diaz, U., Quldlello, R.D., 1990. Definite influence of location and climatic conditions on the fatty acid composition of sunflower seed oil. JAOCS 67: 618–623.
- Mantese, A.I., Medan, D., Hall, A.J., 2006. Achene structure, development and lipid accumulation in sunflower cultivars differing in oil content at maturity. Annals of Botany London 97: 999–1010.
- Martínez-Rivas, J.M., Sperling, P., Lühs, W., Heinz, E., 2001. Spatial and temporal regulation of three different microsomal oleate desaturase genes (FAD2) from normal-type and high-oleic varieties of sunflower (Helianthus annuus L.). Molecular Breeding 8: 159–168.
- Miller, J.F., Zimmerman, D.C., Vick, B.A., 1987. Genetic control of high oleic acid content in sunflower oil. Crop Science 27: 923–926.
- Möllers, C., Schierholt, A., 2002. Genetic variation of palmitate and oil content in a winter oilseed rape doubled haploid population segregating for oleate content. Crop Science 42: 379–384.
- R Development Core Team, 2012. R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria, ISBN 3-900051-07-0.
- Rebetzke, G., Pantalone, V., Burton, J., Carver, B.F.,Wilson, R., 1996. Phenotypic variation for saturated fatty acid content in soybean. Euphytica 91: 289–295.
- Robertson, J.A., Green, V.E., 1981. Effect of planting date on sunflower seed oil content, fatty acid composition and yield in Florida. Journal of the American Oil Chemists' Society 58: 698–701.
- Roche, J., Bouniols, A., Mouloungui, Z., Barranco, T., Cerny, M., 2006. Management of environmental crop conditions to produce useful sunflower oil components. European Journal of Lipid Science and Technology 108: 287–297.
- Roche, J., Essahat, A., Bouniols, A., Asri, M.E., Mouloungui, Z., Mondiès, M., Alghoum, M., 2004. Diversified composition of sunflower (Helianthus annuus L.) seeds within cultural practices and genotypes (Hybrids and Populations). Helia 27: 73–98.
- Rondanini, D., Savin, R., Hall, A.J., 2003. Dynamics of fruit growth and oil quality of sunflower (Helianthus annuus L.) exposed to brief intervals of high temperature during grain filling. Field Crops Research 83: 79–90.
- Schneiter, A.A., Miller, J.F., 1981. Description of sunflower growth stages. Crop Science 21: 901–903.
- Soldatov, K.I., 1976. Chemical mutagenesis in sunflower breeding. In: Proceedings of the 7th Intern. Sunf. Conf. Krasnodar, pp. 352–357.
- Triboï-Blondel, A., Bonnemoy, B., Falcimagne, R., Martignac, M., Messaoud, J., Philippon, J., Vear, F., 2000. The effect of temperature from flowering to maturity on seed composition of high oleic sunflower inbreeds and mid oleic hybrids. In: Proceedings of the 15th Intern. Sunf. Conf. Toulouse, pp. A67–A72.
- Van der Merwe, R., Arno, H., Herselman, L., Labuschagne, M., 2012. Physicochemical and oxidative stability characteristics of high- and mid-oleic sunflower seed oil. In: Proceedings of the 18th Intern. Sunf. Conf. Mar del Plata, pp. 955–960.
- Varès, D., Lacombe, S., Griveau, Y., Bervillé, A., Kaan, F., 2002. Inheritance of oleic acid content of F1 seed in a complete diallel Cross between seven sunflower lines. Helia 25: 105–112.
- Velasco, L., Peréz-Vich, B., Fernández-Martinez, J.M., 2000. Inheritance of oleic acid content under controlled environment. In Proceedings of the 15th Intern. Sunf. Conf. Toulouse, pp. 31–36.
- Velasco, L., Peréz-Vich, B., Fernández-Martinez, J.M., 2007. Relationships between seed oil content and fatty acid composition in high stearic acid sunflower. Plant Breeding 126: 503–508.
- Vick, B.A., Jan, C.C., Miller, J.F., 2004. Two-year study on the inheritance of reduced saturated fatty acid content in sunflower seed. Helia 27: 25–40.
- Were, B.A., Onkware, A.O., Gudu, S., Welander, M., Carlsson, A.S., 2006. Seed oil content and fatty acid composition in East African sesame (Sesamum indicum L.) accessions evaluated over 3 years. Field Crops Research 97: 254–260.
- Zheljazkov, V.D., Vick, B.A., Baldwin, B.S., Buehring, N., Astatkie, T., Johnson, B., 2009. Oil content and saturated fatty acids in sunflower as a function of planting date, nitrogen rate, and hybrid. Agronomy Journal 101: 1003–1111.
- Zuil, S., Izquierdo, N., Lujánd, J., Cantarero, M., Aguirrezábal, L.A.N., 2012. Oil quality of maize and soybean genotypes with increased oleic acid percentage as affected by intercepted solar radiation and temperature. Field Crops Research 127: 203–214.